

<b>BACHELOR OF SCIENCE (BOTANY)</b>		
Programme/Class: <i>Bachelor of Science</i>	Year: <b>III</b>	Semester: <b>V</b> <b>Paper-I</b>
Subject: BOTANY		
Course Code: B040501T	Course Title: <b>Plant Physiology, Metabolism &amp; Biochemistry</b>	
<b>Course outcomes:</b>		
After the completion of the course the students will be able to:		
1. Understand the role of Physiological and metabolic processes for plant growth and development.		
2. Learn the symptoms of Mineral Deficiency in crops and their management.		
3. Assimilate Knowledge about Biochemical constitution of plant diversity.		
4. Know the role of plants in development of natural products, nutraceuticals, dietary supplements, antioxidants		
Credits: <b>4</b>	<b>Core Compulsory</b>	
Max. Marks: <b>25+75</b>	Min. Passing Marks:	
Total No. of Lectures-Tutorials-Practical (in hours per week) <b>4-0-0</b>		
Unit	Topic	No. of Lectures(60hrs)
<b>I</b>	<b>Plant water relation, Mineral Nutrition, Transpiration and translocation in phloem</b> Importance of water, water potential and its components; Transpiration and its significance; Factors affecting transpiration; Root pressure and guttation. Criteria of essentiality of elements; Role of essential elements; Symptoms of mineral deficiency in major crops, Transport of ions across cell membrane, active and passive transport, Composition of phloem sap, girdling experiment; Pressure flow model.	7
<b>II</b>	<b>Carbon Oxidation</b> Krebs cycle, Glycolysis, fate of pyruvate- aerobic and anaerobic respiration and fermentation, regulation of glycolysis, oxidative pentose phosphate pathway, oxidative decarboxylation of pyruvate, regulation of Krebs cycle, mitochondrial electron transport, oxidative phosphorylation, ATP-Synthetase, Chemiosmotic mechanism, P/O ratio, cyanide-resistant respiration, factors affecting respiration.	7
<b>III</b>	<b>Nitrogen Metabolism</b> Nitrate assimilation, biological nitrogen fixation (examples of legumes and non-legumes), Physiology and biochemistry of nitrogen fixation, Ammonia assimilation (GS-GOGAT), reductive amination and transamination, amino acid synthesis.	8
<b>IV</b>	<b>Lipid Metabolism &amp; Photosynthesis</b> Lipid Metabolism: Synthesis and breakdown of triglycerides, -oxidation, glyoxylate cycle, gluconeogenesis and its role in mobilization of lipids during seed germination, -oxidation. ; Photosynthesis: Pigments, Action spectra and Enhancement effect, Electron transport system and Photophosphorylation, C3 & C4 photosynthesis, CAM- Reaction and Significance	7
<b>V</b>	<b>Plant Development, Movements, Dormancy &amp; Responses</b> Developmental roles of Phytohormones (auxins, gibberellins, cytokinins, ABA, ethylene.) autonomic & paratonic movements, Control and Coordination in plants, Photoperiodism (SDP, LDP, Day neutral plants); Phytochrome (discovery and structure), red and far red-light responses on photomorphogenesis, Seed physiology & Dormancy, Vernalization & Senescence	8



**This course can be opted as an elective by the students of following subjects:** Open to all but special for following: B.Sc. Math, B.Sc. Statistics, B.Sc. Nutrition, B.Sc. Biophysics, B.Sc. Biotech,

**Suggested Continuous Evaluation Methods:** Continuous Internal Evaluation shall be based on allotted Assignment and Class Tests

Internal Assessment	Marks
Class Interaction	5
Quiz	5
Seminar	7
Assignment (Charts/ Flora/ Rural Service/ Technology Dissemination)	8
	25

**Course prerequisites:**

**Qualification:** To study this course, a student must have qualified 10+2 with Biology/ NSQF level 3 from Sector Skill Councils / Diploma holder from ITI in (Biology/ Agriculture/ Forestry/ Biotech/ /Gardening)

**Facilities: Smart and Interactive Class**

**Other Requisites: Video collection, Books, CDs, Access to On-line resources, Display Charts**

**Suggested equivalent online courses:**

<https://www.classcentral.com/course/swayam-plant-physiology-and-metabolism-17732>

<https://www.wiziq.com/course/3249-plant-physiology-in-10-live-online-classes>

<https://www.easybiologyclass.com/plant-physiology-free-lecture-notes-online-tutorials-lecture-notes-ppts-mcqs/>

[https://onlinecourses.swayam2.ac.in/cec19\\_bt09/preview](https://onlinecourses.swayam2.ac.in/cec19_bt09/preview)

Programme/Class: <b>Bachelor of Science</b>	Year: <b>III</b>	Semester: <b>V Paper-II</b>
Subject: <b>BOTANY</b>		
Course Code: <b>B040502T</b>	Course Title: <b>Molecular Biology &amp; Bioinformatics</b>	
<b>Course outcomes:</b>		
After the completion of the course the students will be able to:		
1. Understand nucleic acids, organization of DNA in prokaryotes and Eukaryotes, DNA replication mechanism, genetic code and transcription process.		
2. Know about Processing and modification of RNA and translation process, function and regulation of expression.		
3. Gain working knowledge of the practical and theoretical concepts of bioinformatics		
Credits: <b>4</b>	<b>CC / Elective</b>	
Max. Marks: <b>25+75</b>	Min. Passing Marks:	
Total No. of Lectures-Tutorials-Practical (in hours per week) <b>4-0-0</b>		
<b>Unit</b>	<b>Topic</b>	<b>No. of Lectures(60hrs)</b>
<b>I</b>	<b>Genetic material</b> Miescher to Watson and Crick- historic perspective, Griffith's and Avery's transformation experiments, Hershey-Chase, bacteriophage experiment, DNA structure, types of DNA, types of genetic material. DNA replication (Prokaryotes and eukaryotes): semi-conservative. DNA replication (Prokaryotes and eukaryotes): bidirectional replication, semi-conservative, semi discontinuous RNA priming, $\theta$ (theta) mode of replication, replication of linear, dsDNA, replicating the 5' end of linear chromosome including replication enzymes.	<b>7</b>



1. Primrose, SB. 1995. Principles of Genome Analysis. Blackwell Science Ltd. Oxford, UK..
2. E.J. Gardner and D.P. Snustad. PRINCIPAL OF GENETICS (1984), John Wiley & Sons, Ney York.
3. Watson, J. D., Baker T.A., Bell, S. P., Gann, A., Levine, M., and Losick, R., 2008 Molecular Biology of the Gene 6th edition. Cold Spring Harbour Lab. Press, Pearson Pub.
4. Freifelder - Molecular Biology.
5. P.K. Gupta. BIOTECHNOLOGY AND GENOMICS. Rastogi Publications, 7th Reprint (1st Edition): 2016-2017.
6. Ghosh, Z., Mallick, B. (2008). Bioinformatics – Principles and Applications, 1st edition. New Delhi, Delhi: Oxford University Press.
7. Baxevanis, A.D. and Ouellette, B.F., John (2005). Bioinformatics: A Practical Guide to the Analysis of Genes and Proteins, 3rd edition. New Jersey, U.S.: Wiley & Sons, Inc.
8. Roy, D. (2009). Bioinformatics, 1st edition. New Delhi, Delhi: Narosa Publishing House.
9. Andreas, D., Baxevanis, B.F., Francis, Ouellette. (2004). Bioinformatics: A practical guide to the analysis of genes and proteins, 3rd edition. New Jersey, U.S.: John Wiley and Sons.
10. Pevsner J. (2009). Bioinformatics and Functional Genomics, 2nd edition. New Jersey, U.S.: Wiley Blackwell.
11. Xiong J. (2006). Essential Bioinformatics, 1st edition. Cambridge, U.K.: Cambridge University Press
12. A Textbook Of Basic And Molecular Genetics (pb) ISBN : 9788188826193 Edition : 01 Year : 2018 Author : Dr. Parihar

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**This course can be opted as an elective by the students of following subjects:**

Open to all but special for following: B.Sc. Math, B.Sc. Statistics, B.Sc. Nutrition, B.Sc. Biophysics, B.Sc. Biotech, B.Sc. Forestry, B.Sc. Agriculture.

**Suggested Continuous Evaluation Methods:** Continuous Internal Evaluation shall be based on allotted Assignment and Class Tests. The marks shall be as follows:

Internal Assessment	Marks
Class Interaction	5
Quiz	5
Seminar	7
Assignment (Charts/ Flora/ Rural Service/ Technology Dissemination)	8
	<b>25</b>

**Course prerequisites:**

**Qualification:** To study this course, a student must have qualified 10+2 with Biology/ NSQF level 3 from Sector Skill Councils / Diploma holder from ITI in (Biology/ Agriculture/ Forestry/ Biotech)

**Facilities: Smart and Interactive Class**

**Other Requisites: Video collection, Books, CDs, Access to On-line resources, Display Charts**

**Suggested equivalent online courses:**

<https://www.edx.org/learn/molecular-biology>

<https://www.vlab.co.in/broad-area-biotechnology-and-biomedical-engineering>

<https://www.classcentral.com/course/swayam-genetic-engineering-theory-and-application-14090>

<https://www.coursera.org/courses?query=genetics>

<https://www.coursera.org/courses?query=molecular%20biology>

<https://www.edx.org/learn/genetic-engineering>

<https://www.mooc-list.com/tags/genetic-engineering>

<https://www.classcentral.com/course/edx-molecular-biology-part-1-dna-replication-and-repair-2907>

<https://nptel.ac.in/courses/102/103/102103013/>

Programme/Class: <i>Bachelor of Science</i>		Year: <b>III</b>	Semester: <b>V</b> <b>Paper-III</b>
Subject: <b>Botany</b>			
Course Code: B040503P		Course Title: <i>Experiments in physiology, Biochemistry &amp; molecular biology</i>	
<b>Course outcomes:</b>			
After the completion of the course the students will be able to:			
<ol style="list-style-type: none"> <li>1. Know and authentic the physiological processes undergoing in plants along with their metabolism</li> <li>2. Identify Mineral deficiencies based on visual symptoms</li> <li>3. Understand and develop skill for conducting molecular experiments for genetic engineering</li> </ol>			
Credits: 2		<b>Core Compulsory</b>	
Max. Marks: <b>25+75</b>		Min. Passing Marks:	
Total No. of Lectures-Tutorials-Practical (in hours per week) <b>0-0-2</b>			
Unit	Topic*		No. of Lectures(60 hrs)
<i>*(Perform any three from each unit based on facility)</i>			
<b>I</b>	<b>Plant water relation, Mineral Nutrition and translocation in phloem</b> 1. Determination of osmotic potential of plant cell sap by plasmolytic method using leaves of Rhoec / Tradescantia. 2. Osmosis – by potato osmoscope experiment 3. Effect of temperature on absorption of water by storage tissue and determination of Q10. 4. Experiment to demonstrate the transpiration phenomenon with the bell jar method 5. Experiment for demonstration of Transpiration by Four-Leaf Experiment: 6. Structure of stomata (dicot & monocot) 7. Determination of rate of transpiration using cobalt chloride method. 8. Experiment to measure the rate of transpiration by using Farmer’s Potometer 9. Experiment to measure the rate of transpiration by using Ganong’s potometer 10. Effect of Temperature on membrane permeability by colorimetric method. 11. Study of mineral deficiency symptoms using plant material/photographs.		<b>8</b>
<b>II</b>	<b>Nitrogen Metabolism, Photo Synthesis &amp; Respiration</b> 1. A basic idea of chromatography: Principle, paper chromatography and column chromatography; demonstration of column chromatography. 2. Separation of plastidial pigments by solvent and paper chromatography. 3. Estimation of total chlorophyll content from different chronologically aged leaves (young, mature and senescence) by Arnon method. 4. Effect of HCO <sub>3</sub> concentration on oxygen evolution during photosynthesis in an aquatic plant and to find out the optimum and toxic concentration (either by volume measurement or bubble counting). 5. Measurement of oxygen uptake by respiring tissue (per g/hr.) 6. Determination of the RQ of germinating seeds. 7. Effect of light intensity on oxygen evolution in photosynthesis using Wilmott’ bubble		<b>8</b>
<b>III</b>	<b>Plant Development, Movements, Dormancy &amp; Responses</b> <ol style="list-style-type: none"> <li>1. Geotropism and phototropism — Klinostat</li> <li>2. Hydrotropism <ol style="list-style-type: none"> <li>a. Measurement of growth — Arc and Liver Auxonometer</li> </ol> </li> <li>3. To study the phenomenon of seed germination (effect of light).</li> <li>4. To study the induction of amylase activity in germinating grains.</li> </ol>		<b>8</b>

	<ol style="list-style-type: none"> <li>5. Test of seed viability by TTC method.</li> <li>6. To study the effect of different concentrations of IAA on <i>Avena</i> coleoptile elongation (IAA bioassay)</li> </ol>	
<b>IV</b>	<p><b>Techniques for biochemical analysis</b></p> <ol style="list-style-type: none"> <li>1. Weighing and Preparation of solutions -percentage, molar &amp; normal solutions, dilution from stock solution etc.</li> <li>2. Separation of amino acids by paper chromatography.</li> <li>3. Detection of organic acids: citric, tartaric, oxalic and malic from laboratory samples.,</li> <li>4. Qualitative Analysis of carbohydrates,</li> <li>5. Estimation of reducing sugar by anthrone method,</li> <li>6. Qualitative Analysis of Lipids</li> <li>7. Qualitative analysis of Amino acids and Proteins</li> <li>8. Quantitative Analysis of Nucleic Acids,</li> <li>9. Analysis of dietary supplements, nutraceuticals &amp; antioxidants</li> <li>10. Testing of adulterants in food items.</li> </ol>	<b>8</b>
<b>V</b>	<p><b>Genetic material</b></p> <ol style="list-style-type: none"> <li>1. Instruments and equipments used in molecular biology.</li> <li>2. Preparation of LB medium and cultivating E.coli on it.</li> <li>3. Isolation of Genomic DNA</li> <li>4. Isolation of DNA from plants</li> <li>5. Examination of the purity of DNA by agarose gel electrophoresis.</li> <li>6. Quantification of DNA by UV-spectrophotometer</li> <li>7. Estimation of DNA by diphenylamine method.</li> </ol>	<b>7</b>
<b>VI</b>	<p><b>Preparation of models/ charts:</b></p> <ol style="list-style-type: none"> <li>1. Study of experiments establishing nucleic acid as genetic material (Avery et al, Griffith's, Hershey &amp; Chase's and Fraenkel &amp; Conrat's experiments)through photographs</li> <li>2. Numericals based on DNA re-association kinetics (melting profiles and Cot curves)</li> <li>3. Study of DNA replication through photographs: Modes of replication - Rolling circle, Theta and semi-discontinuous ; Semiconservative model of replication (Messelson and Stahl's experiment); Telomerase assisted end-replication of linear DNA</li> <li>4. Study of structures of : tRNA (2D and 3D); prokaryotic RNA polymerase and eukaryotic RNA polymerase II through photographs</li> <li>5. Study of the following through photographs: Assembly of Spliceosome machinery; Splicing mechanism in group I &amp; group II introns; Ribozymes and Alternative splicing</li> <li>6. Understanding the regulation of lactose (lac) operon (positive &amp; negative regulation) and tryptophan (trp) operon (Repression and De-repression &amp; Attenuation) through photographs.</li> <li>7. Understanding the mechanism of RNAi by photographs</li> </ol>	<b>7</b>
<b>VII</b>	<p><b>Genetic Engineering</b></p> <ol style="list-style-type: none"> <li>1. Isolation of protoplasts.</li> <li>2. Construction of restriction map of circular and linear DNA from the data provided.</li> <li>3. Isolation of plasmid DNA.</li> <li>4. Restriction digestion and gel electrophoresis of plasmid DNA (demonstration/ photograph).</li> <li>5. Calculate the percentage similarity between different cultivars of a species using RAPD profile. Construct a dendrogram and interpret results.</li> </ol>	<b>7</b>





**HiMedia Biotechnology & Molecular biology Kits/Chemicals, Micropipettes, Elisa reader/Microtitre Reader**

**Suggested equivalent online courses:**

<https://www.edx.org/learn/molecular-biology>

<https://krishikosh.egranth.ac.in/handle/1/5810039999>

<https://www.classcentral.com/course/swayam-genetic-engineering-theory-and-application-14090>

<https://www.coursera.org/courses?query=genetics>

<https://www.coursera.org/courses?query=molecular%20biology>

<https://www.edx.org/learn/genetic-engineering>

<https://www.mooc-list.com/tags/genetic-engineering>

<https://www.classcentral.com/course/edx-molecular-biology-part-1-dna-replication-and-repair-2907>

Programme/Class: <b>Bachelor of Science</b>	Year: <b>III</b>	<b>Semester: V Paper-IV</b>
Subject: <b>BOTANY</b>		
Course Code: - B040504R	Course Title: <b>Project in Botany for Pre-graduation</b>	
<b>Course outcomes:</b>		
<ul style="list-style-type: none"> <li>● Project work will supplement field experimental learning and deviations from classroom and laboratory transactions.</li> <li>● project work will enhance the capability to apply gained knowledge and understanding for selecting, solving and decision-making processes.</li> <li>● It will promote creativity and the spirit of enquiry in learners.</li> <li>● They will learn to consult Scientists, libraries, laboratories and herbariums and learn importance of discussions, Botanical &amp; field trips, print and electronic media, internet etc. along with data documentation, compilation, analysis &amp; representation in form of dissertation writing.</li> <li>● It will enhance their abilities, enthusiasm, and interest.</li> </ul>		
Credits: <b>03</b>	Core: <b>Compulsory</b>	
Max. Marks: <b>25+75</b>	Min. Passing Marks: <b>.....</b>	
Total No. of Lectures-Tutorials-Practical (in hours per week): <b>0-0-3.</b>		
<b>Suggestive List Of PROJECTS</b>		
<ol style="list-style-type: none"> <li>1. Rural Areas: Flora of a city/ village, Biodiversity of Village, Soil &amp; seed testing service provision to farmers,</li> <li>2. Industrial waste management</li> <li>3. water pollution status of rural water &amp; promotion of WASH in villages</li> <li>4. Plant Disease identification in farms, nurseries and orchards.</li> <li>5. Digital portal for plants: Campus, city or particular area</li> <li>6. Rare and endangered plants &amp; their conservation &amp; domestication</li> <li>7. Air pollution tolerance index (APTI) : Screening of sensitive/tolerant plant species at various locations in particular area</li> <li>8. Science Communication by Creating science documentaries of innovators , Internet Science ( Social media, Websites, Blogs, Youtube, Podcast etc.)</li> <li>9. Science Outreach Talks and Public Sensitization for plant biodiversity conservation sensitization of public.</li> <li>10. Phytochemistry of medicinal plants &amp; their antimicrobial, nutraceutical and antioxidant properties</li> <li>11. Study of pollen grains in different flowers</li> <li>12. Study of stomata in different plants</li> <li>13. Study of various types of secretory and special tissues in plants.</li> </ol>		
Refer: libraries, journals, Memoirs, encyclopaedias, herbaria, Museums, etc.		
This course can be opted as an elective by the students of following subjects:		<b>Open to all</b>
<b>Suggested Continuous Evaluation Methods:</b>		
Continuous Internal Evaluation shall be based on allotted Assignment and Class Tests. The marks shall be as follows:		
<b>Internal Assessment</b>	<b>Marks</b>	
Class Interaction	<b>5</b>	
Seminar	<b>10</b>	
Thesis/dissertation	<b>10</b>	
	<b>25</b>	
<b>Course prerequisites:</b>		

**Qualification:** To study this course, a student must have qualified 10+2 with Biology/ NSQF level 3 from Sector Skill Councils / Diploma holder from ITI in (Biology/ Agriculture/ Biotech/ Forestry/ Microbiology/Gardening /biomedical Science.

**Facilities:** Smart and Interactive Class

**Other Requisites:** All listed under all papers of the course.

**Suggested equivalent online courses:**

<https://ndl.iitkgp.ac.in/>

[https://asiafoundation.org/what-we-do/books-for-asia?gclid=CjwKCAiA7939BRBMEiwA-hX5J-QhBITSyPnvj3r8ycio-L9f5uTy1a6oEoALCLa9Ebu0pyz858yQZxoC5wkQAvD\\_BwE](https://asiafoundation.org/what-we-do/books-for-asia?gclid=CjwKCAiA7939BRBMEiwA-hX5J-QhBITSyPnvj3r8ycio-L9f5uTy1a6oEoALCLa9Ebu0pyz858yQZxoC5wkQAvD_BwE)

<http://www.dli.ernet.in/>

<http://www.ulib.org/>

<http://www.tkdl.res.in/>

<http://www.vigyanprasar.gov.in/digilib>

Directory of Open Access Repositories (DOAR)<http://www.opendoar.org>

Registry of Open Access Repositories (ROAR)<http://roar.eprints.org/>

[http://www.iscnagpur.ac.in/knowledge\\_learning\\_files/5.7\\_General\\_Open\\_Access\\_e-Resources.pdf](http://www.iscnagpur.ac.in/knowledge_learning_files/5.7_General_Open_Access_e-Resources.pdf)